EPIGENETICS:
THE DOGMA-DEFYING DISCOVERY THAT GENES LEARN FROM EXPERIENCE.

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Introduction

Epigenetics challenges our acceptance of the dichotomy between nature and nurture. It is coming to be understood that nurture somehow writes its way into our genes; that experiences, or the effects of experiences on gene transcription, may continue throughout our own lives and into those of the next generation. For anyone engaged in the challenge of helping clients who have suffered traumatic, neglectful or abusive experiences, or for anyone offering clients a new experience through the psychotherapeutic relationship, an understanding of epigenetics is pertinent and illuminating. Here I introduce the concept of epigenetics, describe some epigenetic mechanisms, and review the findings of a number of recent studies that seek to understand why, what, and how our genes can learn from experience.
The engaging debate of whether it is nature or nurture that has the most influence over who we are and what kind of lives we lead often illuminates the attitudes the debaters hold with regard to their own lives. In my early twenties, I tended to champion the influence of the environment, as although I felt affected by childhood adversity, I did not like to think of myself as passively, fatalistically accepting nature—my genetic inheritance—as my lot in life. Discoveries in biology have made this debate and my wrangling with it entirely passé with the enthralling field of epigenetics, which allows for degrees of gene expression or “shades of grey” that release us from the “black and white” dichotomy of genetic determinism. Epigenetics provides insights into how the environment dynamically impacts on our gene expression as humans, influencing, amongst other traits, our and our children’s health, ability to learn and remember, and responses to stress. The fabric of the lives led by our parents and grandparents, from their diet (Katada, Imhof, & Sassone-Corsi, 2012) to their education, the care they received, traumas they may have suffered, and perhaps many other experiences leave their legacy written alongside our DNA as “instructions for interpretation.” The purpose of this as yet inappreciably sophisticated and elegant code of epigenetic information, if it can be properly be called a “purpose,” is to record, use, and pass on information about the intricacies of the environment experienced in both our lives and those that have gone before us, so that we and our children can be as well prepared for survival in our particular environment as it possible to be. Life, over lives, continually perceives the environment, and the epigenetic instructions for interpreting the DNA are written, used, revised, and may be inherited, along with our genes.

Epigenetics is the contemporary study of how the environment influences gene expression both within and, through heritable changes in DNA, beyond the lifetime of an organism. The idea that organisms can inherit environmentally acquired characteristics is, however, the old idea of Lamarckian inheritance. In 1809, Jean-Baptiste Lamarck suggested that an organism would acquire traits through adapting to its environment, and that these traits would then be inherited by its offspring (Handel & Ramagopalan, 2010). Lamarck’s theory was overlooked in favor of Darwin’s natural selection theory of evolution, as the two explanations appeared at the time to be mutually exclusive, but the advent of epigenetics has made it possible for these theories to be reconciled. “Epigenetics” literally means “above the genes,” and is the means by which the environment “marks” the genes, dramatically or subtly, changing their level of expression either transiently or for our lifetime, or, through inheritability, throughout our children’s and grandchildren’s lifetimes. A formal definition of epigenetic events proposed by Adrian Bird is: “the structural adaption of chromosomal regions so as to register, signal or perpetuate altered activity states” (Bird, 2007, p. 398). This definition encompasses the broad remit of epigenetic marks from transient, where the epigenetic mark ascribed by the environmental adaption lasts only a few hours, to heritable, where the environmental effects last over a generation. The brain-derived neurotrophic factor (BDNF) gene, implicated in psychiatric disorders and learning and memory (Autry & Monteggia, 2012; Boulet et al., 2012; Lu, Christian, & Lu, 2008; Minichiello, 2009; Musumeci & Minichiello, 2011), is subject to both short- and long-term epigenetic marking in rodents. For example, following the favorable social experience of being reared in a communal nest, mice challenged with one hour in a mildly stressful novel environment generate hippocampal BDNF faster than mice raised in a standard nest, as a result of an epigenetic mark on the BDNF gene (Branchi, Karpova, D’Andrea, Castren, & Alleva, 2011). However, rat pups subjected to a rat equivalent of childhood maltreatment are epigenetically marked by this experience, reducing the level of BDNF in their pre-frontal cortex throughout their adult life. The offspring of these rats also carry the same epigenetic mark on their BDNF gene even when they have been cross-fostered to non-maltreating mothers (Roth, Lubin, Funk, & Sweatt, 2009). Thus, epigenetics, the mechanisms by which our genes record or adapt to the environment, can shape gene expression over a few minutes, an hour, or a lifetime, and can even shape the gene expression pattern of the next generation. It is even possible for genes to “remember” an event and make a contingency plan for its recurrence, as in the case of the corticotropin-releasing hormone gene of rat pups. In response to maternal deprivation, the promoter region of this gene is epigenetically marked. Later, following a stressful experience in adulthood, the pre-recorded epigenetic mark leads to a hypersensitive stress response, observed as a more actively transcribed corticotropin-releasing hormone gene and increased levels of the stress hormone corticosterone (Chen et al., 2012).

There are three main types of epigenetic modification: DNA methylation and histone modification, outlined here together with examples of these mechanisms in action, and translational regulation by micro RNA (not pertinent to this review, but see (Haramati et al., 2011; Sato, Tsuchiya, Meltzer, & Shimizu, 2011))
for useful introductions).

Types of epigenetic modification, Level I: DNA methylation

Methylation involves the addition of a methyl group to mammalian DNA, and can occur in response to environmental influence, making a stable, potentially heritable addition to the DNA that enhances or represses the transcription of a gene. Methylation is not a mutation, as the sequence of bases (adenine, guanine, cytosine, and thymine) remains the same. The methyl group is added to a cytosine nucleotide, usually followed by a guanine nucleotide (a CpG site), by enzymes known as DNA methyltransferases (DNMTs). The promoter regions or transcriptional start site of genes (the point at which RNA polymerase II begins transcription of the DNA into mRNA) are frequently rich in CpG sites, and methylation of these regions is associated with long-term silencing of genes. Methylation of CpG sites within the body of the gene is more ambiguous and context specific and can lead to either repression or activation (Jones, 2012). Methylation of DNA can also be bound by Methyl Binding Domain (MBD) proteins such as MeCP2 which recruit other protein complexes to re-model the local chromatin (DNA + histone packaging), leading either to repression or activation of specific genes. MeCP2 is an important point of integration between types of epigenetic modification, as MeCP2 binds methylated DNA but also can recruit chromatin re-modeling proteins which implement the other major type of epigenetic alteration, histone modification, described below (Cedar & Bergman, 2012). Methylation of DNA is reversible but instead of de-methylation occurring via the action of a single enzyme, it occurs through a multistep process. It is intriguing that almost half of the methylated DNA in the brain is 5-hydroxymethylcytosine, an intermediate formed during the multistep de-methylation of DNA (Szulwach et al., 2011), a process which is dependent on neuronal activity (Guo, Su, Zhong, Ming, & Song, 2011). The contemporary view of DNA methylation is that, in the brain especially, it is a highly dynamic process, responsive to neural activity (and therefore experience) and crucial for plasticity-related functions such as learning, memory and behavioral adaptation (Baker-Andresen, Ratnu, & Bredy, 2013).

Types of epigenetic modification, Level II: Histone modification

The DNA in the nucleus of each eukaryotic cell, including in the case of humans, contains all of the genetic material. So a human skin cell carries the same complete genetic code as a neuron or a parietal cell of the stomach. Each specialized cell need only be able to transcribe a small fraction of the total DNA it carries in order to perform its specific function. Since the rest of its DNA will never need to be accessed, it can be packed away as efficiently as possible. Histone proteins facilitate this efficient packaging of DNA, largely due to the electrostatic attraction between the negatively charged DNA and the positively charged histone proteins. DNA is coiled around a core octamer of histone proteins, like thread on a spool, forming a single nucleosome. Multiple nucleosomes together are known as chromatin, which has different conformations reflecting the accessibility of the genes for transcription. Euchromatin is open chromatin, conceivable as separate beads on a string; here the genes are accessible, whereas in condensed chromatin or heterochromatin, the genes are inaccessible and un-transcribed, as they are so densely packed (Jenuwein & Allis, 2001).

Like most proteins, the histone proteins comprising the core around which the DNA winds itself can have chemical groups attached to them that may either subtly change their charge, and thus how tightly they are bound to the DNA, or change their affinity for other protein-binding partners. In this way, histone modifications alter the chromatin structure and hence the accessibility of genes, but can also affect the binding of other molecules including DNA methyltransferases—the enzymes that add methyl groups to DNA, not the histone proteins that the DNA coils around (Ooi et al., 2007). This interaction forms another point of integration between DNA-modifying and histone-modifying epigenetic mechanisms. His-
tones may be acetylated, methylated (including bi- or tri-methylated), phosphorylated or ubiquitinylated on specific residues, and each group or combination of groups may signal a precise outcome. For example, an epigenetic modification of a tri-methylated lysine4 residue on histone3 signals an actively transcribed gene, whereas a tri-methylation on lysine27 of histone3 signals a repressed gene state. If lysines 4 and 27 are both tri-methylated, the gene is thought to be “poised,” ready for activation (Bernstein et al., 2006). As there are four histones (two of each make up the octamer core), and at least four chemical groups that can attach to multiple residues, there are a large number of possible combinations of epigenetic marks that can be made on a histone octamer, giving rise to the concept of a “histone code” that can nuance gene expression (Jenuwein & Allis, 2001). There are enzymes that catalyze the addition and removal of each type of chemical group. Histone acetylases (HATs), for example, add acetyl groups to the histone, reducing the electrostatic charge difference and therefore the affinity between the histone and the DNA, thus loosening the coil. Accordingly, HATs tend to be associated with gene activation, whereas histone de-acetylases (HDACs) remove the acetyl group, increase the electrostatic attraction, tighten the coil, and are thus considered transcriptional repressors (Bannister & Kouzarides, 2011).

In contrast to the epigenetic modification of a methylation made to DNA, histone modifications are easily reversible and highly dynamic, although the enzymes that catalyze the addition and removal of various groups are themselves regulated. The histone code also has great flexibility in determining how long a gene’s activity may be enhanced or repressed, as epigenetic marks may record an experience on a gene without necessarily changing its level of transcription. They may transiently enhance or repress gene expression, or may recruit (or repel) DNA methylases that can administer de novo methylations on the DNA which result in the gene becoming stably repressed throughout the life of the organism and in the next generation (Ooi, et al., 2007). It is interesting to note that Valproic acid, a known teratogen used in the treatment of bipolar disorder, is a histone de-acetylase inhibitor but also leads to reduced levels of DNA methylation, demonstrating that a transient and easily reversible epigenetic histone acetylation is frequently followed by a more stable and less easily reversed DNA methylation, silencing the gene (Alonso-Aperte, Ubeda, Achon, Perez-Miguelsanz, & Varela-Moreiras, 1999; Gottlicher, 2004). Thus, histone modification and DNA methylation, whilst distinct epigenetic mechanisms, can act in synergy to produce permanent gene silencing.

The epigenetic impact of early maternal care

Many animal studies of epigenetics have looked at the impact of early maternal care on the hypothalamic-pituitary-adrenal (HPA) axis or, put more simply, the stress axis. This axis co-ordinates the response to stress from the brain, which perceives the stress, to the adrenal gland, which releases glucocorticoids such as cortisol (corticosterone in rodents) in response to the stressor. The hypothalamus receives neural input—our perception of the stress—and releases corticotropin-releasing hormone (CRH) that acts on the anterior pituitary gland to release adrenocorticotropic hormone (ACTH). This in turn acts on the cortex of the adrenal gland to release cortisol. The multiple steps in this process allow multiple opportunities for the glucocorticoids to negatively feed back at several levels of the HPA axis, including the hippocampus and the hypothalamus, limiting CRH release, and at the pituitary, limiting ACTH release (Anacker, Zunszain, Carvalho, & Pariante, 2011; Pariante & Lightman, 2008). Glucocorticoids feed back via glucocorticoid receptors. More receptors means a swifter, more efficient negative feedback
mechanism resulting in lower levels of circulating CRH, ACTH and cortisol, and consequently an HPA axis that readily returns to homeostasis. However, in response to chronically high levels of circulating glucocorticoids, glucocorticoid receptors are down-regulated; as a result, the negative feedback mechanism is less efficient, and the HPA axis is slower to return to homeostasis or may become dysregulated (Anacker, et al., 2011; Sapolsky, Meaney, & McEwen, 1985). The HPA axis is an adaptive system subject to early-life epigenetic programming both prenatally and postnatally through variations in maternal care, separation and abuse.

Early pre-natal stress in mice has been epigenetically linked to enduring changes in HPA axis reactivity and a depressive phenotype in male offspring. Male adult mice whose mothers had been mildly stressed early in utero had higher levels of CRH and fewer glucocorticoid receptors in their hippocampus, the former increasing the activation of the HPA axis and the latter reducing the ability to regulate the HPA axis. These mice exhibited behaviors associated with depression and, following exposure to restraint stress, had increased levels of corticosterone compared to mice whose mothers had not been subjected to early pre-natal stress. When the promoter regions of relevant genes were examined, it was discovered that the promoter for corticotropin-releasing hormone was hypo-methylated, increasing the expression of CRH, the hormonal activator of the HPA axis. The glucocorticoid receptor promoter was also hyper-methylated, reducing its level of expression and thereby the mouse’s ability to negatively regulate its HPA axis after activation by CRF. This would at least contribute to, if not cause, the observed depressed behavior (Mueller & Bale, 2008).

Early post-natal stress has also been linked to enduring epigenetic changes that alter the reactivity of the HPA axis. Rat pups that receive enhanced licking and grooming by their mothers in the first ten days of life acquire a permanent increase in their number of glucocorticoid receptors, meaning that their HPA axis is more readily negatively regulated. Thus, when these pups become adults and are exposed to an acute stressor, they release corticosterone, which negatively feeds back through a greater number of glucocorticoid receptors, quickly reducing the levels of CRH, circulating ACTH, and corticosterone (Liu et al., 1997). If this effect was correlated with measurable changes to the epigenome, i.e., DNA methylation or histone code changes at the site of a relevant gene, it would support the hypothesis that maternal care leads to epigenetic changes that moderate the activity of the stress axis. In support of this hypothesis, a landmark study by Weaver’s group (Weaver, Diorio, Seckl, Szyf, & Meaney, 2004) compared rat pups that had either high or low levels of licking and grooming from mothers with whom they were cross-fostered, and found epigenetic differences in the promoter region of the glucocorticoid receptor gene. Pups that had received high levels of licking and grooming had hypo-methylated DNA and hyper-acetylated histone3 at the promoter region of the glucocorticoid receptor gene where a transcription factor binds, leading to enhanced transcriptional activity of the glucocorticoid receptor gene and therefore to more glucocorticoid expression.
receptors. As adults, pups that had received low levels of licking and grooming had fewer glucocorticoid receptors and correspondingly higher levels of circulating corticosterone following restraint stress. Taking the study a step further towards identifying maternal care as a cause of epigenetic changes that modify the reactivity of the HPA axis, a histone de-acetylase (HDAC) inhibitor was administered to the rats which not only enhanced acetylation of the glucocorticoid promoter in the low licking and grooming group, it also removed all of the other observed differences in this group: the glucocorticoid promoter became hypo-methylated, more glucocorticoid receptors were expressed, and the corticosterone response following restraint stress was normalized to the same level as the high licking and grooming group. Essentially, the epigenetic differences brought about by lower levels of maternal care were reversed by the HDAC inhibitor (Weaver et al., 2004). In a follow-up study by the same group (Weaver et al., 2005), rats were infused with a methyl group donor. Even if the rats had received high levels of licking and grooming by their mothers when they were pups, leading to hypo-methylation of the glucocorticoid promoters, if they received the methylating drug as adults, all of the beneficial effects of this extra licking and grooming were reversed; the glucocorticoid promoter became hyper-methylated and fewer glucocorticoid receptors were expressed, leading to higher levels of corticosterone following a stressful experience and higher scores in a test of behavioral responses to stress (Weaver et al., 2005). From their combined studies, the authors primarily concluded that maternal behavior programs the stress response through epigenetic modification of the glucocorticoid promoter, and secondarily concluded that although maternal care makes stable changes to the epigenome, these are pharmacologically modifiable even in adults (Weaver, et al., 2005). Since then, in the case of studies showing that differences in estrogen receptor levels correlated with the level of maternal care received as a pup and the level of maternal care given in turn to offspring (Champagne, Weaver, Diorio, Sharma, & Meaney, 2003), the relationship has also been found to be mediated through epigenetic changes to the estrogen receptor promoter. Thus, receiving a high level of maternal care leads to reduced methylation of the CpG region within the promoter of the estrogen receptor, correspondingly increased levels of expressed estrogen receptors, and the capacity to give a high level of maternal care to the following generation of pups (Champagne et al., 2006).

Separation from mother in infancy has long been known to have emotionally and psychologically detrimental sequelae, and the work of two brilliant and influential researchers, Harry Harlow and John Bowlby, is eloquently discussed in van der Horst and van der Veer’s review (van der Horst & van der Veer, 2008). In the field of epigenetics, the effects of maternal separation on the reactivity of the HPA axis are beginning to be discovered. In the HPA axis, stressful stimuli perceived at the level of the hypothalamus lead to secretion of CRH-stimulating ACTH. Arginine vasopressin (AVP) is also secreted by the hypothalamus, and potentiates the effect of CRH at the anterior pituitary, stimulating more ACTH release and increased HPA activity. Mice that were separated from their mothers for three hours a day for the first ten days of their lives had higher basal levels of corticosterone as well as higher levels following a stressful experience, and also scored higher on behavioral measures of stress and lower on memory tasks. Higher levels of expressed AVP correlated with hypo-methylated DNA in the regulatory region of the AVP gene (Murgatroyd et al., 2009). The methyl-binding protein MeCP2, which binds methylated DNA, ordinarily represses AVP transcription. But in mice that have been separated from their mothers, MeCP2 cannot bind as readily to the AVP promoter, as it is hypo-methylated even when the mouse reaches adulthood, leading to reduced repression by MeCP2 and consequent higher levels of AVP transcription. This results in a persistent hormonal activation of the HPA axis throughout the life of the mouse (Murgatroyd, et al., 2009).

As well as enduringly activating the HPA axis, early life stress induced by maternal separation has been shown to enhance performance in stress-related memory tasks in young rats (Suri et al., 2012). The rats’ improved performance correlated with increased neurogenesis, reduced repressive histone methylation of a BDNF gene promoter, and corresponding-
ly higher levels of BDNF in the hippocampus, compared to controls. Interestingly, once the maternally separated rats reached middle age, all these beneficial effects were lost, although this could be ameliorated by long-term antidepressant treatment. The authors of this study suggested that maternal separation stress induces biologically adaptive responses including epigenetic changes that increase the rat pups’ chances of having to survive alone, without the mother’s help and protection, but that these adaptive changes exact a heavy toll when the rat reaches middle age (Suri et al., 2012). From an evolutionary point of view, as long as the adaptive response to maternal separation allows the rat to survive on its own long enough to be able to reproduce, the exacted toll is worthwhile.

Paradigms that mimic abusive behavior in humans have also been used to examine the epigenomic effects of maternal abuse in rodents, demonstrating that abuse leads to significant hypo-methylation of the BDNF promoter in the pre-frontal cortex. Notably, rats that had suffered maltreatment in infancy also maltreated their own (or cross-fostered) pups, and the effect of their own maltreatment could be seen in their biological offspring as a hypo-methylated BDNF promoter region even when the pups were fostered to a caring mother (Roth et al., 2009). Maltreatment, or abuse, appears to be such a profound environmental influence that its effects linger in the generation following that which experienced the maltreatment firsthand. A similar observation of the trans-generational transmittance of a profound experience has been made by Rachel Yehuda, a researcher into post-traumatic stress disorder (PTSD) who found a higher than normal prevalence of PTSD in the children of mothers who had PTSD as a result of the Holocaust, suggesting a mechanism other than traditional genetic contribution in play, with epigenetics being the obvious candidate (Yehuda, Bell, Bierer, & Schmeidler, 2008). Although specific epigenetic modifications have not yet been linked to experiences that lead to the development of PTSD in humans, a recent study has identified patterns of genes that are differentially methylated in human suffers of PTSD (Uddin et al., 2010). Additionally, in the rat model of PTSD, where the animals are exposed on two occasions to a cat, as well as having a new rat cage mate daily for 31 days, dramatic epigenetic changes have been observed in the promoter region of the BDNF gene in the hippocampus (Roth, Zoladz, Sweatt, & Diamond, 2011). Taken together, these results suggest that the development of human PTSD may occur through traumatic events causing epigenetic changes to specific genes, which, as well as leading to PTSD in the trauma victim, could confer an increased risk of the same in the victim’s children, as the epigenetic signatures on the genes caused by the trauma could be inherited (Yehuda & Bierer, 2009). Although speculative, it seems likely that the inheritance of genes that have been epigenetically marked by trauma could be biologically adaptive and enhance the chance of survival if the environment remains life-threatening for the next generation. Hyper-vigilance and extreme alertness following trauma, whilst profoundly exhausting and distressing, may be life-saving in a dangerous environment.

Human studies looking at the epigenetic impact of the environment

Gathering data for human studies tends to involve the use of postmortem tissue and detailed retrospective analyses of significant life events or trauma. A recent study using these methodologies compared the differences in DNA methylation across the whole genome of 25 men with a history of severe child abuse compared to 16 controls, and found that fewer genes are actively transcribed in men who suffered abusive childhoods. The methylation profiles of the men showed 248 genes were hyper-methylated and 114 genes were hypo-methylated in the group with a history of abuse compared to the control group (Labonte et al., 2012). Of these, the top five most hyper-methylated genes were neuronal, and many of the genes that were differently methylated in the group who had experienced childhood abuse were related to plasticity, i.e., genes that are known to have a role in learning or adaptive mechanisms. A similar study using blood samples found that children raised in institutions had more methylated, and thus fewer expressed, genes than children raised by their biological parents (Naumova et al., 2012). It is tempting to speculate that perhaps lower levels of care restrict the variety of genes that can be expressed, whereas higher levels of care are the epigenetic gateway to our available genome. We speak of wanting to give our children “every opportunity”, but the reality of what epigenetic opportunities are available to our children could largely depend on the care we give them very early in life. Access to the widest potential of their genome could be profoundly influenced by their experience of our care, and if the genes that are accessed by enhanced care perform plasticity-related functions, the effect of that care is compounded for good or ill. Poor care may mean that fewer plasticity-related genes are expressed, thus restricting a child (and the adult’s) potential to learn, remember or adapt within their environment, manifesting in less flexibility and perhaps more stereotyped or rigid responses.
Studies that identify the epigenetic effects of levels of care or experiences on specific genes have been undertaken in humans. In a striking human parallel to Weaver's (2004) rodent study showing that low levels of maternal care epigenetically modified the glucocorticoid receptor promoter by hyper-methylation, reducing the transcription of the glucocorticoid receptor gene, McGowan et al. (2009) found that suicide victims who had a history of childhood abuse had hyper-methylated glucocorticoid promoters and fewer HPA axis-regulating hippocampal glucocorticoid receptors than controls. Both prenatal stress and even the birth experience have also been linked to epigenetic changes. The depressive mood of mothers in the third trimester of pregnancy has been linked to hyper-methylated glucocorticoid receptor promoters and higher cortisol responses in three-month-old babies (Oberlander et al., 2008), while infants born by Caesarean section have significantly higher levels of DNA methylation in their leucocytes compared to vaginal births at the time of delivery, although this normalizes after 3–5 days (Schлинzig, Johansson, Gunnar, Ekstrom, & Norman, 2009).

**Pharmacotherapeutics, enriching environments, and reversible epigenetic change**

Pharmacological treatments can reverse epigenetic marks caused by environmental experiences on both the BDNF gene (Roth, et al., 2009; Suri, et al., 2012) and the glucocorticoid receptor gene (Weaver, et al., 2005; Weaver, Meaney, & Szyf, 2006). Antidepressant treatment has frequently been shown to restore levels of BDNF in animal models of depression (Balaratanasingam & Janca, 2012), and in the social defeat, mouse model of depression, imipramine has been demonstrated to act through epigenetic mechanisms, reversing the down-regulating effect that social defeat has on BDNF transcription levels, although it does so by a compensatory HDAC-inhibiting mechanism rather than direct reversal of repressive histone methylations (N. M. Tsankova et al., 2006). The HDAC inhibitor, valproic acid, a treatment in certain psychiatric disorders, also epigenetically enhances BDNF transcription, facilitating the forgetting (or extinction) of fear-conditioned learning (Bredy et al., 2007; Whittle et al., 2013). Other histone-modifying agents may become useful psychoactive medications. It has recently been demonstrated that the acetylating agent L-acetyl-carnitine, already available as a dietary supplement, enhances transcription of the type II metabotropic glutamate receptor, which has a swift and enduring anti-depressant effect in rodents (Nasca et al., 2013). As epigenetic mechanisms mediate the effects of environment, the involvement of epigenetic mechanisms in psychiatric disorders that have an environmental component is axiomatic. Whilst pharmacotherapeutics holds promise for the treatment of psychiatric disorders (Grayson, Kundakovic, & Sharma, 2010; N. Tsankova, Renthal, Kumar, & Nestler, 2007), for the many people who fall below the threshold of a psychiatric diagnosis or who prefer non-medicating treatments, the very nature of the epigenetic mechanism suggests an alternative to drug treatment in the form of new, desirable environmental experiences to overwrite previous negative or undesirable experiences written on the epigenome. Environmental enrichment is proof of the principle that new environmental experiences can reverse or ameliorate the epigenetic effects of a previously impoverished or stressful environment.

Enriching environments provide opportunities for enhanced sensory, motor, cognitive, and social stimulation, and have a multitude of beneficial effects, from enhancing neural plasticity and learning and memory, to conferring resilience to depression and ameliorating the effects of many brain disorders including Alzheimer’s and Huntington’s diseases and stroke (Bekinschtein, Oomen, Saksida, & Bussey, 2011; D’Andrea, Gracci, Alleva, & Branchi, 2010; Nithianantharajah & Hannan, 2006). The effects of an enriched environment in rodents are profound, and can even mitigate the effects of prenatal stress or maternal separation on the HPA axis (Francis, Diorio, Plotsky, & Meaney, 2002; Morley-Fletcher, Rea, Maccari, & Laviole, 2003). Analogously to the effects of trauma, the effects of enrichment can be seen in the next generation, improving learning and memory (Arai, Li, Hartley, & Feig, 2009) and, following the socially enriching experience of being reared in a communal nest, enhancing levels of maternal care in the next generation, reducing anxiety behavior, and increasing litter sizes (Curley, Davidson, Bateson, & Champagne, 2009). It is likely that epigenetic mechanisms mediate the benefits of enrichment in terms of both the generation experiencing the enrichment and the trans-generational effects of enrichment that are observed. Recent work shows that rats whose parents had enriching experiences have less methylated DNA than rats whose parents experienced standard housing (Mychasiuk et al., 2012), suggesting that enrichment (such as high levels of care) increases the number of genes that will be actively transcribed. This perhaps reflects a need for greater biological complexity in order to be able to thrive in a more complex (enriched) environment. One gene that is extremely responsive to the environment is BDNF, which is epigenetically modified by the
experience of abuse (see above) as well as by the experience of an enriched environment. A recent study demonstrated increased levels of permissive histone methylations, enhancing transcription of the gene and raising levels of hippocampal BDNF in mice that experienced a month in an enriched environment (Kuzumaki et al., 2011). The role of BDNF in learning and memory behaviors, combined with its epigenetically regulated transcription, suggests this gene is a critical transducer of the experienced environment. Environmental enrichment is a powerful tool to effect epigenomic changes, which subsequently affect the levels of BDNF and modulate HPA axis activity, which, in turn, builds resilience to, or delays the onset of, various psychiatric or neurodegenerative disorders.

The relevance of epigenetics for psychotherapy

When I learned that real-life subjective experiences could regulate gene transcription, it changed the way I understood myself. I was not just an immutable collection of transcribed genes. My genes had my experiences of early care, my traumas and dramas and education written onto them in the tiniest molecular inscription of the histone code and DNA methylations. Furthermore, even as I was born, the most salient facts of my parents’ and even their parents’ lives—the kinds of care they had received, their education, their sense of safety or fear—was etched into my genes too, to help me adapt to what was biologically anticipated to be a similar environment. I did not know about epigenetics when I underwent psychotherapy, but if I had, it may have inspired my curiosity about my parents’ lives and informed my developing narrative that who they were, who I was, was a more intricate and elegantly entwined interplay between our genes and our environments than I could ever have imagined. Neither they nor I was responsible or to blame; instead, we were placed to offer an enriched experience of a caring relationship, albeit an asymmetrical one, where one can receive empathic, attuned attention that may be wholly novel and life changing. By offering a new, benevolent experience of relationship, psychotherapy can be reasonably assumed to change gene expression, calming a hyper-reactive stress axis and ameliorating the effects of an epigenetically scarred BDNF promoter, and in so doing, changing our responses to stress and perhaps our capacity to learn and flexibly respond to emotionally challenging situations. For the client in psychotherapy, as in many situations in life, it really is the experience that counts.

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